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Interkalibracija metod vrednotenja ekološkega stanja -
fitobentos

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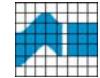
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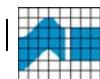
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POVZETEK

Interkalibracija metod vrednotenja ekološkega stanja z biološkimi elementi kakovosti je obligacija vsake države članice Evropske unije v skladu z Vodno direktivo. V letu 2012 se je zaključila druga faza ineterkalibracije. Glede na preliminarne rezultate interkalibracije in komentarje »peer-review« smo dopolnili mejne vrednosti za vrednotenje ekološkega stanja velikih rek na podlagi fitobentosa in podali obrazložitev, zakaj je Trofični indeks primeren tudi za vrednotenje ekološkega stanja jezer na podlagi fitobentosa, čeprav je bil razvit za vrednotenja ekološkega stanja rek. Ker smo poročila oz. znastvene obrazložitve pripravili za Evropsko komisijo, so pripravljena v angleščini.



1 PYHTOBENTHOS-BASED ASSESSMENT OF THE ECOLOGICAL STATUS OF LARGE RIVERS IN SLOVENIA

1.1 Introduction

Large rivers in Slovenia are defined as rivers with a catchment area $>2500 \text{ km}^2$ and/or mean annual discharge $>50 \text{ m}^3/\text{s}$ (Urbanič 2011). However, in the "large river" intercalibration group only data from very large rivers (water-bodies with a catchment area $>10000 \text{ km}^2$) were considered.

1.2 Official name of the assessment method

Metodologija vrednotenja ekološkega stanja rek s fitobentosom in makrofiti v Sloveniji; fitobentos (Ecological status assessment system for rivers using phytobenthos and macrophytes in Slovenia; Phytobenthos)

1.3 Assessment system

Ecological status assessment system for rivers using phytobenthos and macrophytes in Slovenia consists of 2 modules:

- Organic pollution: Saprobic index (Zelinka & Marvan 1961, Rott et al. 1997) based on diatom data
- Eutrophication: Trophic index (Rott et al. 1999) based on diatom data and RMI (Kuhar et al. 2011) based on macrophytes (RMI is not always applicable)

Final classification of the water body is obtained using worst case approach of both modules (Fig. 1).

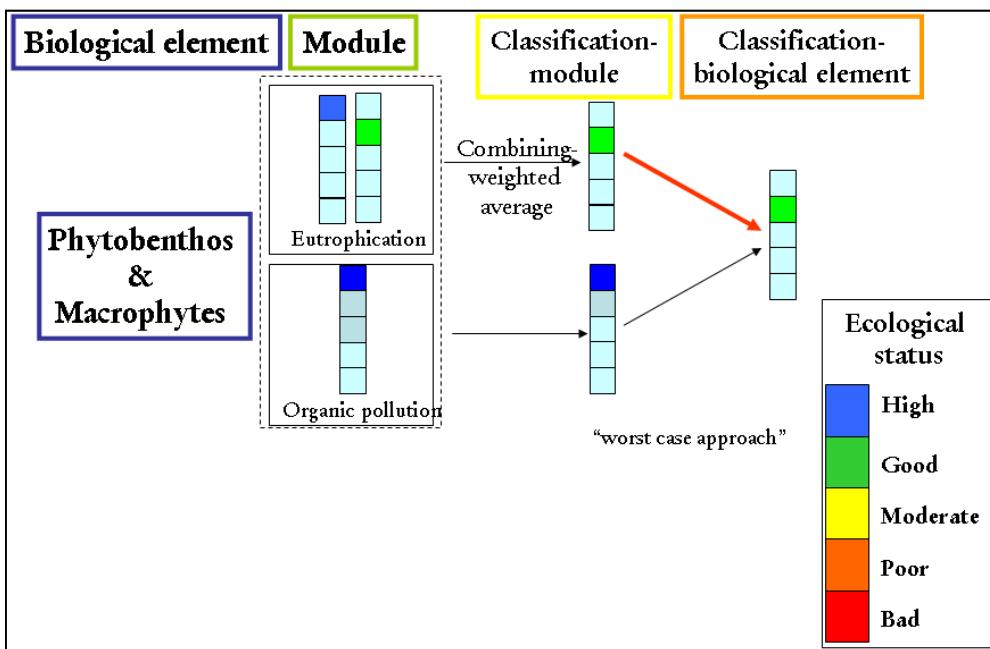


Figure 1. Phytobenthos and macrophytes-based classification of Slovenian rivers.

For the intercalibration purpose of phytobenthos-based assessment methods and ecological quality boundary values EQRs were defined not including a macrophyte metric (RMI index). Thus, EQR was obtained calculating minimum of transformed EQRs of Trophic index and Saprobič index (Tabs 2-4).

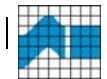
1.4 Reference conditions

Reference values of both indices (Trophic and Saprobič index) for large rivers were defined using regression equations between log total phosphorous values and indices. Reference values were calculated as index values at $\log \text{TP} = -2$ ($\text{TP} = 10 \mu\text{g/L}$).

Ecological class boundary setting

Boundary setting

- a) High/Good and Good/Moderate boundaries were derived from metric variability at stressor class 1.



- b) Other boundaries (Moderate/Poor and Poor/Bad) by equidistant division.

Boundary description

- a) High/Good boundary was defined as 10th percentile value of the pressure class 1 (out of three).
- b) Good/Moderate boundary was defined as 50th percentile value of the pressure class 1 (out of three).
- c) Other boundary values (Moderate/Poor and Poor/Bad) were defined using equidistant division of the remaining EQR gradient.
- d) Lower anchor was defined as highest possible index value based on the Slovenian operational taxa list; it is not maximum index value.

Boundary community

Expressed by specific metric values (Trophic index and Saprobični index), no verbal description.

1.5 Ecological classification

Original boundary values of the Saprobični and Trophic Index describing ecological classes are given in the Table 1.

Table 1. Boundary values and normalised boundary values of the Saprobični index and Trophic index.

Boundary value	SI	SI_EQR	TI	TI_EQR
Reference value	1.74	1	1.96	1
High/Good	1.87	0.94	2.39	0.78
Good/Moderate	2.02	0.86	2.94	0.49
Moderate/Poor	2.61	0.58	3.26	0.33
Poor/Bad	3.21	0.29	3.58	0.16
Lower anchor	3.80	0	3.90	0



In order to combine EQR values in the final BQE based EQR and to compare EQR values with other Biological quality elements EQR values, all values were piecewise linearly transformed and five equidistant classes were obtained (Tables 2-4).

Table 2. Piecewise linear transformation equations for Saprobič Index (SI).

SI_EQR	SI_EQR_transformed
>0.94	$0.8+0.2*(SI_EQR-0.95)/(0.05)$
0.94-0.86	$0.6+0.2*(SI_EQR-0.86)/(0.08)$
0.85-0.58	$0.4+0.2*(SI_EQR-0.58)/(0.27)$
0.57-0.29	$0.2+0.2*(SI_EQR-0.29)/(0.28)$
<0.29	$0.2*(SI_EQR)/(0.28)$

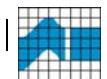
Table 3. Piecewise linear transformation equations for Trophic Index (TI).

TI_EQR	TI_EQR_transformed
>0.78	$0.8+0.2*(TI_EQR-0.79)/(0.21)$
0.78-0.49	$0.6+0.2*(TI_EQR-0.49)/(0.29)$
0.48-0.33	$0.4+0.2*(TI_EQR-0.33)/(0.15)$
0.32-0.16	$0.2+0.2*(TI_EQR-0.16)/(0.16)$
<0.16	$0.2*(TI_EQR)/(0.15)$

Table 4. Transformed boundary values between five ecological status classes of the Ecological status assessment system for rivers using phytobenthos and macrophytes in Slovenia; phytobenthos (PhB_transformed).

Boundary	PhB_transforme
High/Good	0,8
Good /Moderate	0,6
Moderate/Poor	0,4
Poor /Bad	0,2

1.6 Pressure-impact relationships – phytobenthos



Based on very large river data, there was a significant relationship between logarithmic value of the total phosphorous and EQR values of the Ecological status assessment system for rivers using phytobenthos and macrophytes in Slovenia with phytobenthos data (Fig. 2).

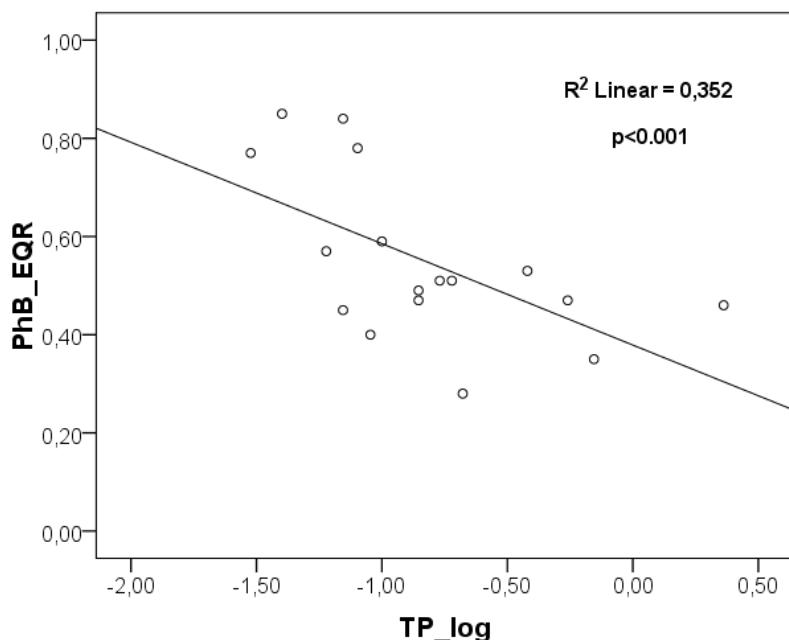


Figure 2. Regression plot of total phosphorous (log transformed data – TP_log) against phytobenthos-based Slovenian national EQR values (PhB_EQR) in large rivers of Slovenia.

1.7 References

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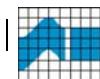


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2 USING (RIVER) TROPHIC INDEX FOR ASSESSMENT OF THE LAKE TROPHIC STATUS IN SLOVENIA

2.1 Introduction

Phytobenthos and macrophytes are one biological element under the Water Framework Directive (Directive 2000/60/ES). In Slovenian lake ecological classification system both sub-elements are used together as one element. Phytobenthos assessment system consists of one metric - Trophic Index (Rott et al. 1998). Trophic Index is calculated as weighted average of the diatom taxa trophic values (TW), where taxa abundance (H), and taxa indicative weights (G) are weighting factors. Individual trophic values (TW) and indicative weights (G) were defined according to the occurrence of the diatom taxa along the eutrophication gradient in rivers (Rott et al. 1998).

The aim of our work is to show that (River) Trophic Index (Rott et al. 1998) can provide a reliable assessment of the trophic status of lakes using lake littoral diatoms.

2.2 Study area

Altogether, 13 lakes were investigated and 96 diatom samples were taken between 2005 and 2011 (Tabs. 1-2).

Table 5. The main characteristics of the studied lakes.

Lake	Ecoregion (Urbanič 2008)	Elevation (m a.s.l.)	Surface area (km ²)	Volume (Mio m ³)	Depth - maximum (m)	Average depth (m)
Blejsko jezero	Alps	475	1.43	26.6	31	19
Bohinjsko jezero	Alps	526	3.28	92.4	45	28
Družmirsko jezero	Alps	360	0.70	25.0	87	24
Velenjsko jezero	Alps	367	1.35	25.0	55	19
Klivnik	Dinaric western Balkan	460	0.36	4.3	20	9
Mola	Dinaric western Balkan	450	0.68	4.3	12	6
Gajševsko jezero	Pannonian lowland	206	0.77	2.6	10	3
Ledavsko jezero	Pannonian lowland	225	2.18	5.7	5	3
Pernica 1	Pannonian lowland	245	0.57	1.2	4	3
Pernica 2	Pannonian lowland	245	0.66	2.1	4	3
Slivniško jezero	Pannonian lowland	292	0.84	4.0	14	5
Šmartinsko jezero	Pannonian lowland	265	1.07	6.5	12	6
Vogeršček	Po lowland	101	0.82	8.5	20	10

**Table 6. Number of sampling sites for each lake and year of sampling.**

Lake/year	2005	2006	2007	2008	2009	2010	2011	Sum
Blejsko jezero	3		7		7	3		20
Bohinjsko jezero	3		7		7	3		20
Družmirsko jezero							3	3
Velenjsko jezero					3		3	6
Klivnik				3				3
Mola				3				3
Gajševsko jezero		3					3	6
Ledavsko jezero		3					3	6
Pernica 1		3						3
Pernica 2		3					3	6
Slivniško jezero					3		3	6
Šmartinsko jezero		5					3	8
Vogerček		3		3				6
Sum	6	20	14	9	20	6	21	96

Physico-chemical parameters and chlorophyll *a* were measured 4 times a year in a vegetation period. Water samples were taken at the deepest part of the lake (Table 3).

Table 7. Minimum and maximum values of measured parameters of the whole dataset and used in the development and validation dataset for Lake Littoral Trophic Index.

Parameter	Dataset Code	Whole		Development		Validation	
		min	max	min	max	min	max
Total Phosphorous – mean (µg/L)	TP-log	3,6	224,0	4,0	224,0	3,6	101,0
Total Nitrogen – mean (µg/L)	TN-log	296	1693	299	1693	296	1534
Secchi depth – mean (m)	Secchi depth	0,3	9,7	0,3	9,0	0,3	9,7
Chlorophyll <i>a</i> – mean (µg/L)	Chlorophyll	1,0	37,6	1,0	36,4	1,0	37,6

2.3 Trophic Index (TI)

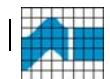
Trophic Index (Rott et al. 1998) is calculated according to the equation:

$$TI = \frac{\sum_{i=1}^n TW_i * G_i * H_i}{\sum_{i=1}^n G_i * H_i}$$

where

TI = Trophic index

TW_i = Trophic value of the taxon »i«



G_i = Indicative weight of the taxon »i«

H_i = Abundance of the taxon »i«

Trophic index shows a response to eutrophication. In lakes of Slovenia we have found a good relationship between mean annual total phosphorous concentrations in lakes and the Trophic Index (Fig. 6).

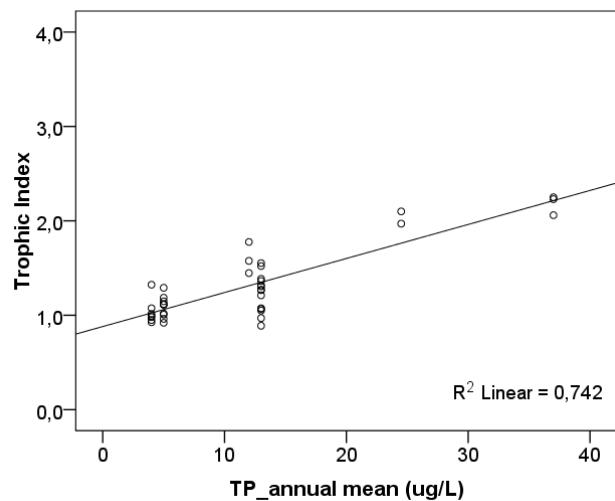


Figure 3. Regression plots of the mean annual Total phosphorous vs. Trophic Index using diatom data from alpine lakes (Slovenian intercalibration dataset).

Comparison of reference and non-reference sites in alpine lakes revealed significant differences in the Trophic Index (Fig. 7).

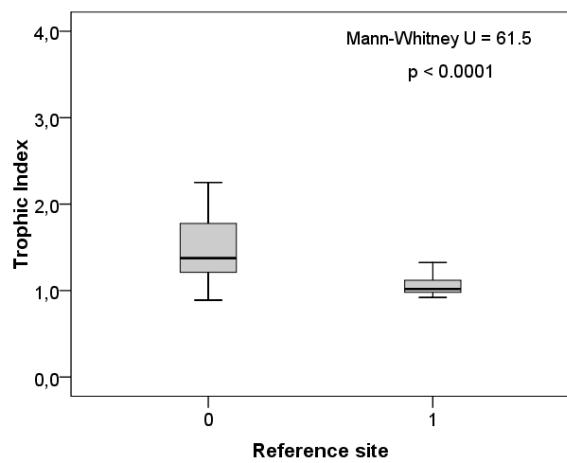
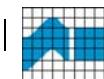


Figure 4. Distribution of Trophic index values between reference (1) and impaired (0) sites of the alpine lakes (Slovenian intercalibration dataset) with the results of the Mann-Whitney U-test.

On average slightly more than 33 diatoma taxa were present in lake littoral samples (Fig. 8). The number of taxa ranged from 10 to 56, whereas in alpine lakes from 25 to 49 (Figs. 9-10). Number of indicator taxa used for the calculation of the Trophic Index ranged from 8 to almost 50, whereas in alpine lakes from 20 to 42 (Figs. 10-11). Percentage of Trophic Index indicator taxa in the diatom sample was always relatively high and on average exceeds 80 % of the present diatom taxa. Only in one diatom sample indicator taxa represent <60% (Fig. 12). In the alpine lake littoral samples percentage of indicator diatom taxa was never below 70% whereas the mean percentage was even slightly higher in comparison to all considered samples (Fig. 13).

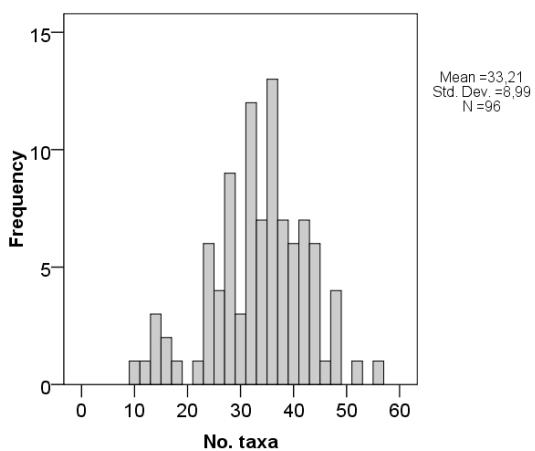
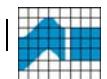


Figure 5. Frequency distribution of number of taxa in lake littoral diatom samples.

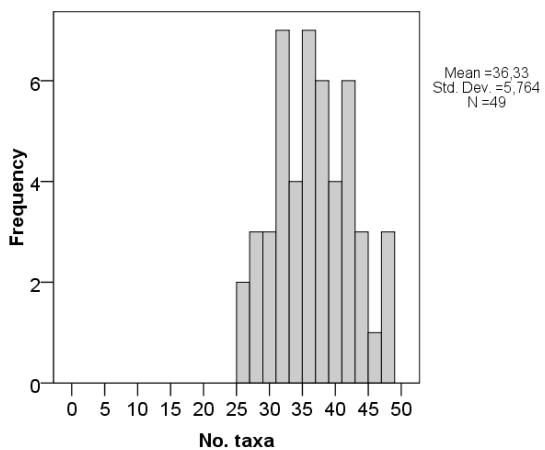
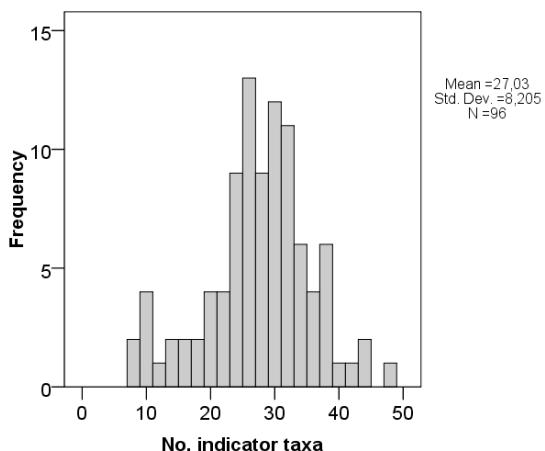


Figure 6. Frequency distribution of number of taxa in alpine lake littoral diatom samples.



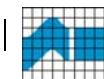


Figure 7. Frequency distribution of number of Trophic Index indicator taxa in lake littoral diatom samples.

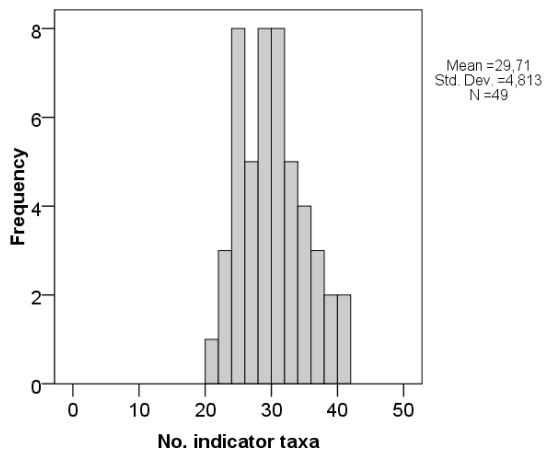


Figure 8. Frequency distribution of number of Trophic Index indicator taxa in alpine lake littoral diatom samples.

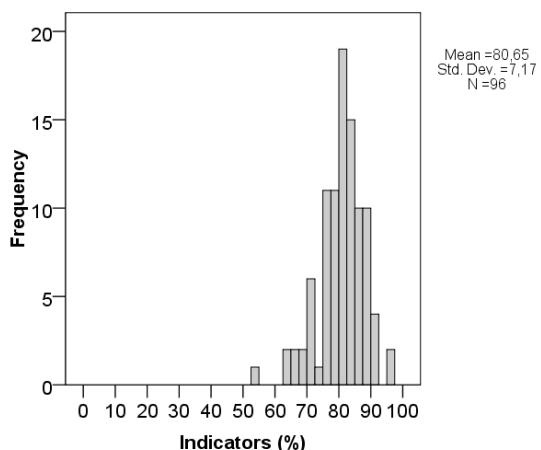


Figure 9. Frequency distribution of percentage of Trophic Index indicator taxa in lake littoral diatom samples.

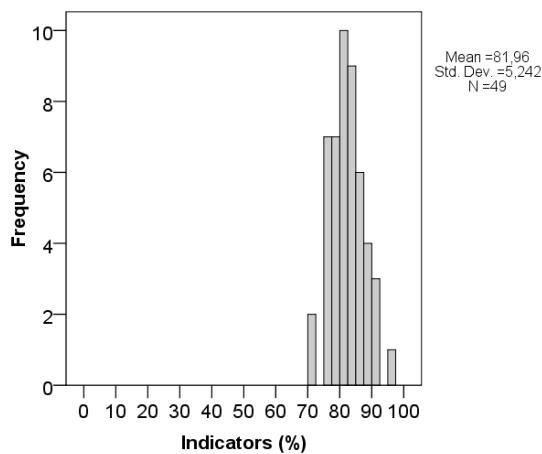
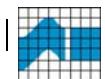


Figure 10. Frequency distribution of percentage of Trophic Index indicator taxa in alpine lake littoral diatom samples.

2.4 Lake Littoral Trophic Index (LLTI)

A whole dataset was divided in a development dataset (62 sites) and a validation dataset (34 sites). A Lake Littoral Trophic Index was developed using a development dataset. A canonical correspondence analysis (CCA) was performed with 185 diatom taxa (Appendix 1) and four environmental parameters (Tables 3-4, Fig. 1).

Table 8. Marginal (Lambda 1) and conditional (Lambda A) effects of the environmental parameters, P-value and F-value.

Variable-code	Lambda 1	Lambda A	P	F
Chlorophyll	0.45	0.45	0.001	6.76
TP-log	0.44	0.24	0.001	3.86
Secchi depth	0.37	0.15	0.001	2.45
TN-log	0.29	0.22	0.001	3.45

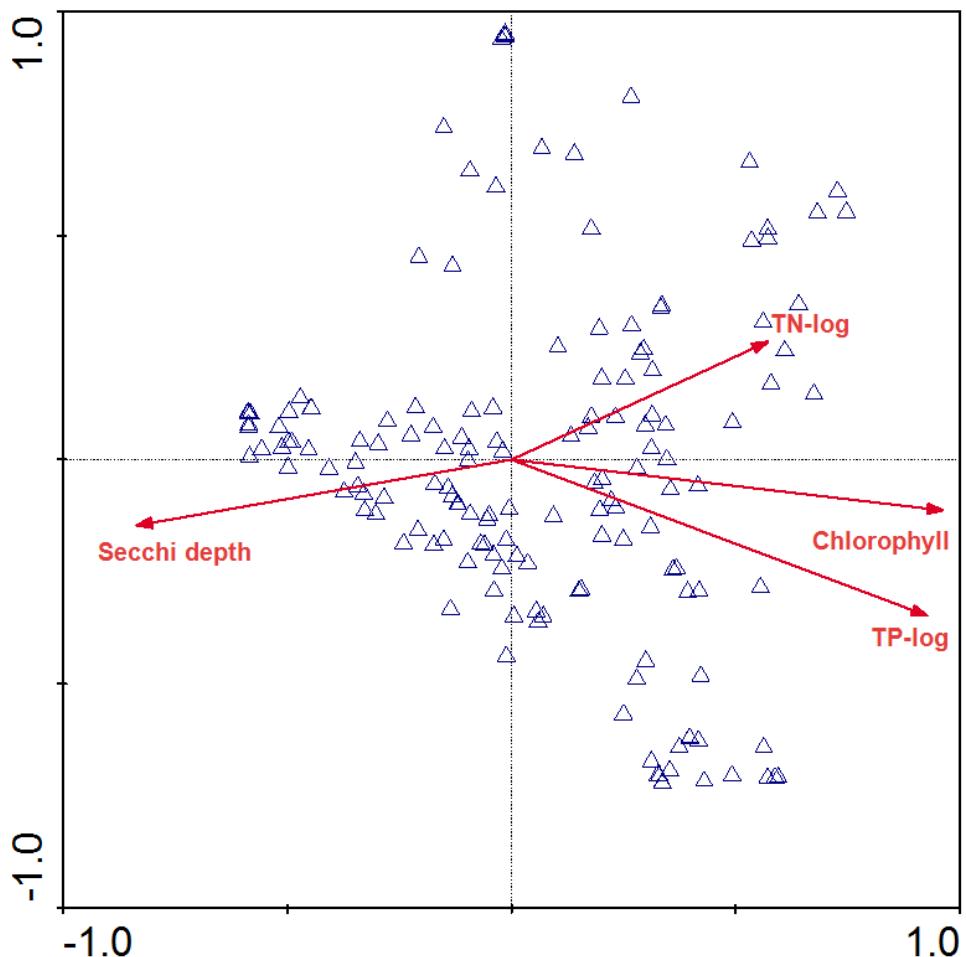
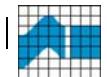


Figure 11. CCA ordination diagram with 185 diatom taxa (open triangles) and four environmental variables (arrows)

The LLTI was developed using trophic preferences (diatom trophic values - Dtv) and tolerance (trophic indicative weights - Tiw) of taxa along the first CCA axes. Diatom trophic values (Dtv) were then determined using CCA ordination axis 1 species scores (biplot scaling):

$$Dtv_i = \frac{SC_{-}CCA1_i}{SC_{-}CCA1_{\max}} \quad ... (1)$$



where SC_CCA1i is the CCA ordination axis 1 species score (biplot scaling) of the i-th taxon and SC_CCA1max is the absolute maximum value of the CCA ordination axis 1 species score (biplot scaling). Trophic indicative weights (Tiw) were determined using the CCA ordination axis 1 species tolerance (root mean squared deviation for species) according to Table 5.

Table 9. Determination of the trophic indicative weight (Tiw) from the CCA axis 1 species tolerance (root mean squared deviation for species).

Tolerance (t_i)	Tiw
$t_i < 0.2$	5
$0.2 < t_i < 0.4$	4
$0.4 < t_i < 0.6$	3
$0.6 < t_i < 0.8$	2
$t_i > 0.8$	1

The LLTI was calculated according to the following equation:

$$LLTI_j = \frac{\sum_{i=1}^n a_i * Dtv_i * Tiw_i}{\sum_{i=1}^n a_i * Tiw_i} \quad ... (2)$$

where a_i is the abundance of the i-th taxon, Dtv_i is the diatom trophic value of the i-th taxon, Tiw_i is the trophic indicative weight of the i-th taxon and n is the number of indicative taxa.

A good relationship was observed between annual mean total phosphorous concentration (log value) and LLTI using development ($R^2 = 0.85$) and validation dataset ($R^2 = 0.70$) (Figs. 2-3). Statistically significant differences were observed in LLTI values between reference sites and impaired sites using data from all lakes (Mann-Whitney U = 71, p <0.0001) and from alpine lakes (Mann-Whitney U = 67, p <0.0001) (Figs. 4-5).

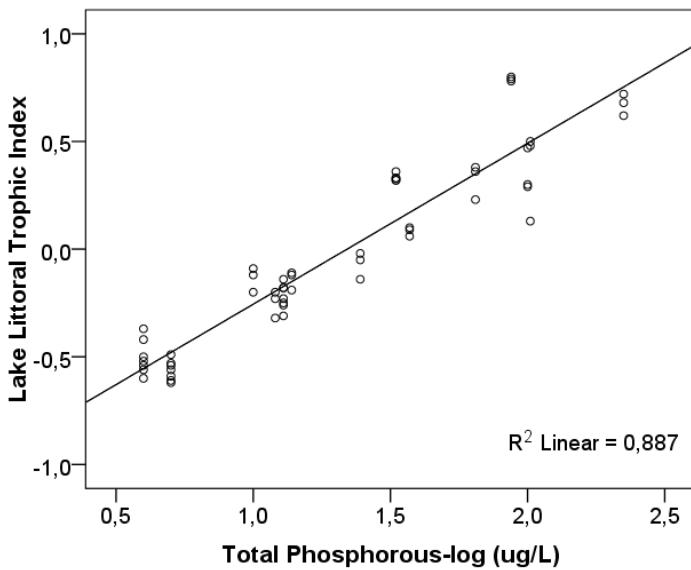
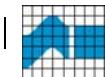


Figure 12. Regression plots of the mean annual Total phosphorous (log value) vs. Lake Littoral Trophic Index using a diatom development dataset

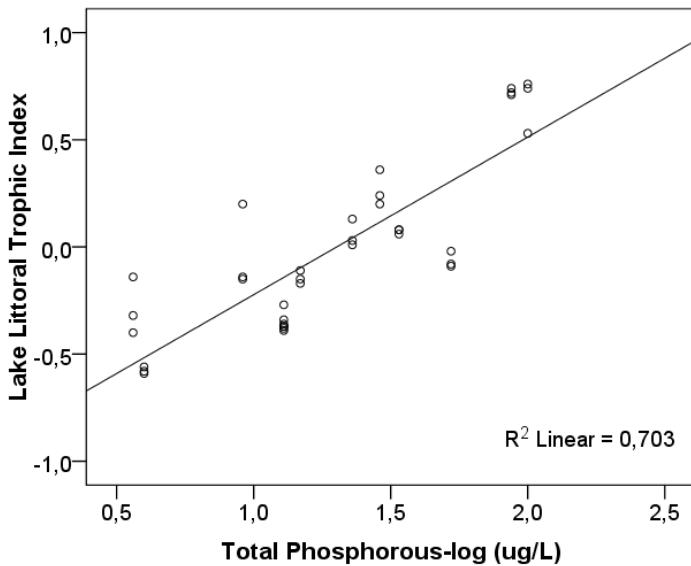


Figure 13. Regression plots of the mean annual Total phosphorous (log value) vs. Lake Littoral Trophic Index using a diatom validation dataset

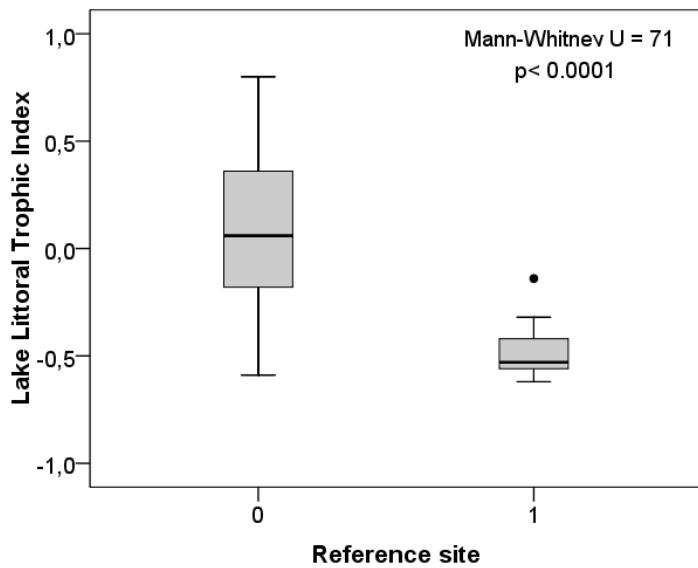


Figure 14. Boxplots of the Lake Littoral Trophic Index values recorded at reference (1) and impaired (0) sites with the results of the Mann-Whitney U-test

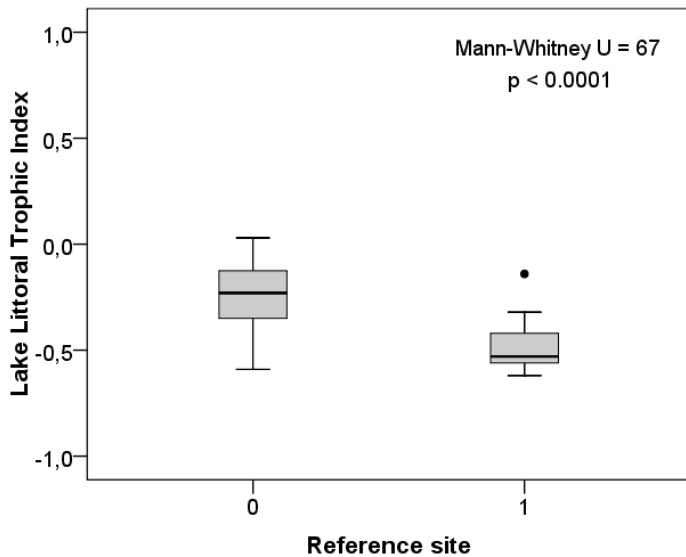
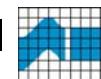


Figure 15. Boxplots of the Lake Littoral Trophic Index (LLTI) values recorded at reference (1) and impaired (0) sites of the alpine lakes with the results of the Mann-Whitney U-test



2.5 Trophic Index (TI) vs. Lake Littoral Trophic Index (LLTI)

A good relationship was observed between TI and LLTI using samples from all lakes ($R^2 = 0.85$) and alpine lakes ($R^2 = 0.74$) (Figs. 14-15).

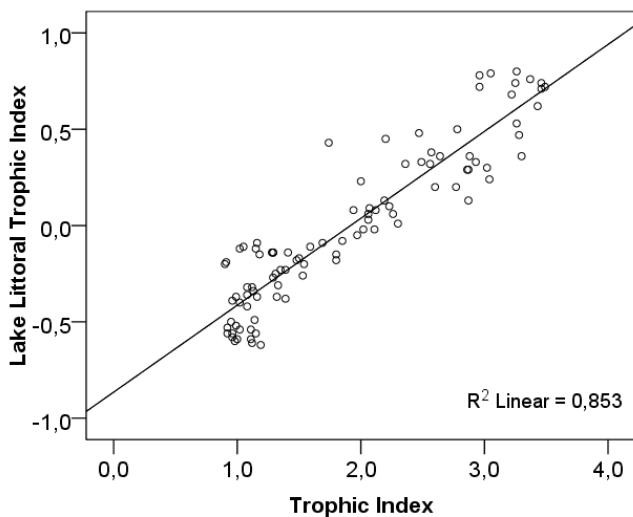


Figure 16. Regression plots of the Trophic Index vs. Lake Littoral Trophic Index using data from all lakes.

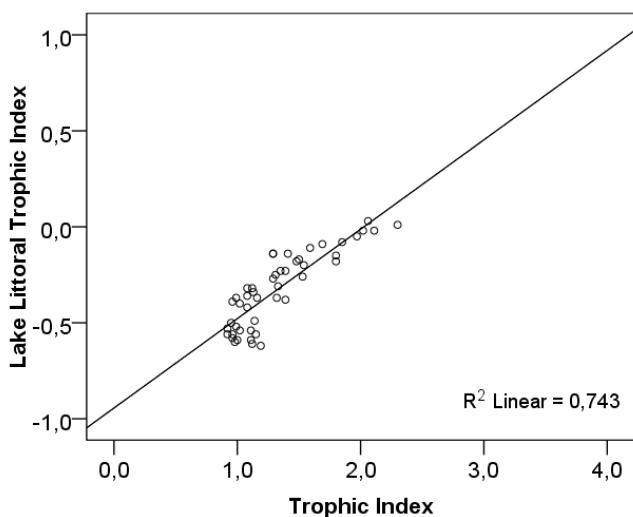
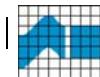


Figure 17. Regression plots of the Trophic Index vs. Lake Littoral Trophic Index using data from alpine lakes.



2.6 Summary

1. (River) Trophic Index (TI) showed good relationship with lake total phosphorous concentrations.
2. Reference sites showed statistically significantly lower TI values than impaired sites.
3. High percentage of diatom taxa occurring in considered littoral samples have assigned trophic values (TW) according to Rott et al. (1998); on average >80% of recorded taxa. In alpine lakes >70% of diatom taxa recorded in each littoral sample were indicator taxa.
4. In the each littoral diatom sample at least eight TI indicator taxa were recorded, whereas on average >25. In samples of alpine lakes at least 20 indicative taxa occurred in the each sample.
5. A relationship between lake littoral diatom taxa and environmental variables representing eutrophication gradient in lakes was tested using Canonical correspondence analysis. Data were collected from varied lake types (lowland and alpine lakes).
6. A new Lake Littoral Trophic Index (LLTI) was developed using littoral diatom data and four environmental variables representing eutrophication gradient.
7. Lake Littoral Trophic Index (LLTI) showed good relationship with mean annual total phosphorous concentration (log data) using development ($R^2 = 0.85$) and validation dataset ($R^2 = 0.70$).
8. Reference sites showed statistically significantly lower LLTI values than impaired sites using all data and alpine data.
9. (River) Trophic Index showed a good relationship with new developed Lake Littoral Trophic Index (LLTI) using samples from all lakes ($R^2 = 0.85$) and alpine lakes ($R^2 = 0.74$).

2.7 Conclusions

(River) Trophic Index (TI) showed a good relationship with the eutrophication gradient. A statistically significant difference in TI was observed between reference and impaired sites and high percentage of recorded littoral diatom taxa was indicative according to TI in all samples. Moreover, a new developed littoral diatom-based trophic index (LLTI) was highly correlated with the (River) Trophic Index using all data and alpine data only. Thus, diatom-based (River) Trophic Index might considerably well address eutrophication pressure in lakes, although lake littoral diatom specific indices might be more applicable.



2.8 References

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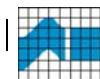
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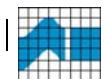
Appendix 1. List of developed diatom trophic values (Dtv) and trophic indicative weights (Tiw) of diatom taxa for calculation of the Lake Littoral Trophic Index (LLTI).

Taxon	Omnidia code	Dtv	Tiw
<i>Achnanthes helvetica</i>	AHAL	-0,44	5
<i>Achnanthes biasolletiana</i>	ABIA	-0,28	5
<i>Achnanthes bioretii</i>	ABIO	-0,78	5
<i>Achnanthes clevei</i>	ACLE	0,49	1
<i>Achnanthes exiqua</i>	AEXG	0,49	1
<i>Achnanthes flexella</i>	AINF	-0,37	5
<i>Achnanthes hungarica</i>	AHUN	0,80	1
<i>Achnanthes lanceolata</i>	ALAN	0,25	3
<i>Achnanthes minutissima v. gracillima</i>	AMGR	-0,65	5
<i>Achnanthes minutissima</i>	AMIN	-0,04	5
<i>Achnanthes oblongella</i>	AOBG	0,09	5
<i>Achnanthes</i> sp.	ACHS	0,42	2
<i>Achnanthes minutissima v.saprophiila</i>	AMSA	0,80	1
<i>Amphora aequalis</i>	AAEQ	-0,02	5
<i>Amphora montana</i>	AMMO	0,39	2
<i>Amphora ovalis</i>	AOVA	0,37	2
<i>Amphora lybica</i>	ALIB	0,34	3
<i>Amphora pediculus</i>	APED	0,13	4
<i>Amphora</i> sp.	AMPS	-0,79	5
<i>Amphipleura pellucida</i>	APEL	-0,12	5
<i>Brachysira vitrea</i>	BVIT	-0,66	5
<i>Anomoeoneis sphaerophora</i>	ASPH	0,45	2
<i>Anomoeoneis vitrea</i>	AVIT	-0,07	5
<i>Asterionella formosa</i>	AFOR	0,19	4
<i>Caloneis alpestris</i>	CAPS	-0,44	5
<i>Caloneis amphisbaena</i>	CAMP	-0,02	5
<i>Caloneis bacillum</i>	CBAC	-0,13	5
<i>Caloneis silicula</i>	CSIL	0,36	2
<i>Fragilaria arcus</i>	FARC	-0,02	5
<i>Cocconeis pediculus</i>	CPED	-0,13	5
<i>Cocconeis placentula</i>	CPLA	0,31	3
<i>Cyclotella meneghiniana</i>	CMEN	0,75	1
<i>Cyclotella ocellata</i>	COCE	0,45	2
<i>Cyclotella</i> sp.	CYLS	0,36	2
<i>Cymatopleura elliptica</i>	CELL	-0,05	5
<i>Cymatopleura solea</i>	CSOL	-0,20	5
<i>Cymbella affinis</i>	CAFF	-0,30	5
<i>Cymbella amphyccephala</i>	CAPH	-0,05	5
<i>Cymbella caespitosa</i>	CCAE	0,02	5
<i>Cymbella cesatii</i>	CCES	-0,69	5
<i>Cymbella cistula</i>	CCIS	0,20	4



Appendix 1. continued

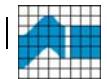
Taxon	Omnidia code	Dtv	Tiw
<i>Cymbella delicatula</i>	CDEL	-0,75	5
<i>Cymbella ehrenbergii</i>	CEHR	0,07	5
<i>Cymbella falaisensis</i>	CFAL	-0,18	5
<i>Cymbella helvetica</i>	CHEL	-0,60	5
<i>Cymbella incerta</i>	CINC	-0,19	5
<i>Cymbella lanceolata</i>	CLAN	-0,01	5
<i>Cymbella microcephala</i>	CMIC	-0,23	5
<i>Cymbella minuta</i>	CMIN	-0,38	5
<i>Cymbella naviculiformis</i>	CNAV	-0,79	5
<i>Cymbella</i> sp.	CYMS	0,78	1
<i>Cymbella prostrata</i>	CPRO	0,33	3
<i>Cymbella silesiaca</i>	CSLE	-0,29	5
<i>Cymbella sinuata</i>	CSIN	-0,61	5
<i>Cymbella tumida</i>	CTUM	0,53	1
<i>Denticula kuetzingii</i>	DKUE	-0,12	5
<i>Denticula tenuis</i>	DTEN	-0,68	5
<i>Diatoma moniliformis</i>	DMON	-0,08	5
<i>Diatoma vulgaris</i>	DVUL	-0,20	5
<i>Diploneis elliptica</i>	DELL	0,24	3
<i>Diploneis oblongella</i>	DOBL	-0,02	5
<i>Diploneis ovalis</i>	DOVA	-0,79	5
<i>Diploneis subconstricta</i>	DSCO	-0,79	5
<i>Epithemia sorex</i>	ESOR	-0,12	5
<i>Epithemia adnata</i>	EADN	-0,54	5
<i>Eunotia arcus</i>	EARC	-0,63	5
<i>Eunotia bilunaris</i>	EBIL	-0,28	5
<i>Fragilaria capucina</i>	FCAP	-0,05	5
<i>Fragilaria capucina</i> v. <i>austriaca</i>	FCAU	-0,47	5
<i>Fragilaria capucina</i> v. <i>capucina</i>	FCAP	-0,23	5
<i>Fragilaria capucina</i> v. <i>distans</i>	FCDI	-0,02	5
<i>Fragilaria construens</i>	FCON	-0,07	5
<i>Fragilaria crotonensis</i>	FCRO	0,26	3
<i>Fragilaria leptostauron</i>	FLEP	-0,78	5
<i>Fragilaria pinnata</i>	FPIN	-0,16	5
<i>Fragilaria capucina</i> v. <i>vaucheriae</i>	FCVA	-0,05	5
<i>Frustulia vulgaris</i>	FVUL	-0,13	5
<i>Gomphonema angustatum</i>	GANG	-0,79	5
<i>Gomphonema augur</i>	GAUG	0,49	1
<i>Gomphonema clavatum</i>	GCLA	-0,50	5
<i>Gomphonema gracile</i>	GGRA	0,31	3
<i>Gomphonema micropus</i>	GMIC	-0,32	5
<i>Gomphonema minutum</i>	GMIN	-0,23	5
<i>Gomphonema pumilum</i>	GPUM	-0,40	5
<i>Gomphonema olivaceum</i>	GOLI	0,21	4

**Appendix 1. continued**

Taxon	Omnidia code	Dtv	Tiw
<i>Gomphonema parvulum</i>	GPAR	0,46	1
<i>Gomphonema</i> sp.	GOMS	-0,09	5
<i>Gomphonema truncatum</i>	GTRU	0,37	2
<i>Gyrosigma acuminatum</i>	GYAC	0,71	1
<i>Gyrosigma attenuatum</i>	GYAT	0,56	1
<i>Gyrosigma nodiferum</i>	GNOD	0,47	1
<i>Gyrosigma scalproides</i>	GSCA	0,80	1
<i>Gyrosigma spencerii</i>	GSPE	0,77	1
<i>Hantzschia amphioxys</i>	HAMP	0,30	3
<i>Aulacoseira granulata</i>	AUGR	0,77	1
<i>Melosira varians</i>	MVAR	0,10	5
<i>Navicula atomus</i>	NATO	0,74	1
<i>Navicula bacillum</i>	NBAC	-0,79	5
<i>Navicula bryophyla</i>	NBRY	-0,78	5
<i>Navicula cari</i>	NCAR	0,08	5
<i>Navicula capitata</i>	NCAP	0,42	2
<i>Navicula cincta</i>	NCIN	0,46	1
<i>Navicula citrus</i>	NCIT	0,90	1
<i>Navicula clementis</i>	NCLE	0,24	3
<i>Navicula contenta</i>	NCON	-0,78	5
<i>Navicula cryptocephala</i>	NCRY	0,42	2
<i>Navicula capitatoradiata</i>	NCPR	0,26	3
<i>Navicula veneta</i>	NVEN	0,42	2
<i>Navicula cuspidata</i>	NCUS	0,50	1
<i>Navicula elginensis</i>	NELG	0,45	2
<i>Navicula gallica</i>	NGAL	-0,78	5
<i>Navicula gallica</i> v. <i>perpusilla</i>	NGPE	-0,78	5
<i>Navicula halophila</i>	NHAL	-0,13	5
<i>Navicula goeppertia</i>	NGOE	0,86	1
<i>Navicula gregaria</i>	NGRE	0,53	1
<i>Navicula lanceolata</i>	NLAN	0,56	1
<i>Navicula menisculus</i>	NMEN	0,18	4
<i>Navicula oblonga</i>	NOBL	-0,02	5
<i>Navicula protracta</i>	NPRO	0,14	4
<i>Navicula pupula</i>	NPUP	0,27	3
<i>Navicula pygmaea</i>	NPYG	0,53	1
<i>Navicula placentula</i>	NPLA	-0,78	5
<i>Navicula radios</i> a	NRAD	-0,15	5
<i>Navicula cryptotenella</i>	NCTE	-0,18	5
<i>Navicula reichardtiana</i>	NRCH	0,01	5
<i>Navicula reinhardtii</i>	NREI	0,45	2
<i>Navicula rhynchocephala</i>	NRHY	-0,78	5
<i>Navicula schroeteri</i>	NSHR	0,75	1

**Appendix 1. continued**

Taxon	Omnidia code	Dtv	Tiw
<i>Navicula</i> sp.	NASP	-0,03	5
<i>Navicula splendicula</i>	NSPD	0,45	2
<i>Navicula subalpina</i>	NSBN	-0,46	5
<i>Navicula trivialis</i>	NTRV	0,39	2
<i>Navicula tripunctata</i>	NTPT	0,56	1
<i>Navicula tuscula</i>	NTUS	-0,66	5
<i>Navicula viridula</i>	NVIR	0,91	1
<i>Navicula viridula v. rostellata</i>	NVRO	0,33	3
<i>Neidium ampliatum</i>	NEAM	0,47	1
<i>Neidium binodis</i>	NBID	-0,02	5
<i>Neidium dubium</i>	NEDU	0,27	3
<i>Nitzschia acicularis</i>	NACI	0,56	1
<i>Nitzschia amphibia</i>	NAMP	0,58	1
<i>Nitzschia angustata</i>	NIAN	-0,67	5
<i>Nitzschia angustatula</i>	NZAG	-0,03	5
<i>Nitzschia constricta</i>	NZCO	0,45	2
<i>Nitzschia capitellata</i>	NCPL	0,05	5
<i>Nitzschia dissipata</i>	NDIS	-0,03	5
<i>Nitzschia dubia</i>	NDUB	-0,13	5
<i>Nitzschia fonticola</i>	NFON	-0,40	5
<i>Nitzschia frustulum</i>	NIFR	0,66	1
<i>Nitzschia gisela</i>	NGIS	-0,78	5
<i>Nitzschia heufleriana</i>	NHEU	0,45	2
<i>Nitzschia incospicua</i>	NINC	0,49	1
<i>Nitzschia levidensis</i>	NLEV	0,79	1
<i>Nitzschia linearis</i>	NLIN	0,42	2
<i>Nitzschia littoralis</i>	NLIT	0,45	2
<i>Nitzschia microcephala</i>	NMIC	-0,20	5
<i>Nitzschia palea</i>	NPAL	0,40	2
<i>Nitzschia paleacea</i>	NPAE	0,76	1
<i>Nitzschia recta</i>	NREC	-0,18	5
<i>Nitzschia sigmoidea</i>	NSIO	0,48	1
<i>Nitzschia sinuata</i>	NSIN	0,27	3
<i>Nitzschia sinuata v. delognei</i>	NSDE	0,49	1
<i>Nitzschia</i> sp.	NZSS	0,23	4
<i>Nitzschia umbonata</i>	NUMB	0,45	2
<i>Nitzschia tryblionella</i>	NTRY	0,82	1
<i>Pinnularia viridis</i>	PVIR	-0,45	5
<i>Rhiocosphenia abbreviata</i>	RABB	0,40	2
<i>Rhopalodia gibba v. minuta</i>	RGMI	-0,79	5
<i>Rhopalodia gibba</i>	RGIB	0,45	2
<i>Stauroneis anceps</i>	STAN	-0,79	5
<i>Stauroneis smithii</i>	SSMI	-0,02	5

**Appendix 1. continued**

Taxon	Omnidia code	Dtv	Tiw
<i>Stephanodiscus</i> sp.	STSP	0,97	1
<i>Surirella angusta</i>	SANG	0,66	1
<i>Surirella bifrons</i>	SBIF	0,44	2
<i>Surirella brebissonii</i>	SBRE	0,45	2
<i>Surirella biseriata</i>	SBIS	-0,02	5
<i>Surirella minuta</i>	SUMI	0,44	2
<i>Surirella ovalis</i>	SOVI	-0,02	5
<i>Surirella tenera</i>	SUTE	1,00	1
<i>Fragilaria ulna v. acus</i>	FUAC	0,72	1
<i>Fragilaria parasitica</i>	FPAR	0,49	1
<i>Fragilaria capucina v.rumpens</i>	FCRP	-0,13	5
<i>Fragilaria ulna</i>	FULN	-0,16	5
<i>Tabellaria flocculosa</i>	TFLO	-0,60	5
<i>Thalassiosira weisflogii</i>	TWEI	0,45	2